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Probing Heteronuclear ¹⁵N-¹⁷O and ¹³C-¹⁷O Connectivities and Proximities by Solid-State NMR Spectroscopy

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Abstract: Heteronuclear solid-state magic-angle spinning (MAS) NMR experiments for probing ¹⁵N-¹⁷O dipolar and J couplings are presented for [²H(NH₃),1-¹³C,¹⁵N,¹⁷O₂]glycine -²HCl and [¹⁵N₂,¹⁷O₂]uracil. Twodimensional ¹⁵N-¹⁷O correlation spectra are obtained using the R³-HMQC experiment; for glycine ²HCl, the intensity of the resolved peaks for the C=O and C-O²H 17 O resonances corresponds to the relative magnitude of the respective ¹⁵N-¹⁷O dipolar couplings. ¹⁷O-¹⁵N REDOR curves are presented for glycine \cdot^{2} HCI; fits of the initial buildup ($\Delta S/S < 0.2$) yield effective dipolar couplings in agreement with (±20%) the root-sum-squared dipolar couplings determined from the crystal structure. Experimental ¹⁵N-¹⁷O REAPDOR curves for the ¹⁵N resonances in glycine ·²HCl and uracil fit well to the universal curve presented by Goldbourt et al. (J. Am. Chem. Soc. 2003, 125, 11194). Heteronuclear ¹³C-¹⁷O and ¹⁵N-¹⁷O J couplings were experimentally determined from fits of the quotient of the integrated intensity obtained in a heteronuclear and a homonuclear spin-echo experiment, $S_{Q}(\tau) = S_{HET}(\tau)/S_{HOM}(\tau)$. For glycine ²HCl, ¹ J_{CO} was determined as 24.7 \pm 0.2 and 25.3 \pm 0.3 Hz for the C=O and C-O²H resonances, respectively, while for uracil, the average of the two NH···O hydrogen-bond-mediated J couplings was determined as 5.1 \pm 0.6 Hz. In addition, two-bond intramolecular J couplings, ${}^{2}J_{OO} = 8.8 \pm 0.9$ Hz and ${}^{2}J_{N1,N3} = 2.7 \pm 0.1$ Hz, were determined for glycine • 2HCI and uracil, respectively. Excellent agreement was found with J couplings calculated using the CASTEP code using geometrically optimized crystal structures for glycine·HCI $[^{1}J_{CO}(C=O) = 24.9 \text{ Hz}, \ ^{1}J_{CO}(C=OH) = 27.5 \text{ Hz}, \ ^{2}J_{OO} = 7.9 \text{ Hz}]$ and uracil $[^{2h}J_{N1,O4} = 6.1 \text{ Hz}, \ ^{2h}J_{N3,O4} = 4.6 \text{ Hz})$ Hz, ${}^{2}J_{N1,N3} = 2.7$ Hz].

1. Introduction

The structural and dynamic information inherent to throughspace dipolar and through-bond J couplings is a key reason why NMR has become such an indispensable analytical tool. Of the elements most commonly found in organic and biological solids (i.e., hydrogen, carbon, nitrogen, and oxygen), heteronuclear solid-state NMR experiments involving the spin I = 1/2 isotopes of hydrogen, carbon, and nitrogen are commonly employed (¹H, ¹³C, and ¹⁵N).¹⁻⁴ The only NMR-active isotope of oxygen is

- Current address: Paul Scherrer Institute, 5232 Villigen PSI, Switzerland.
- ^v Current address: CEA Saclay, SCM, Gif-sur-Yvette 91191, France. (1) Goldbourt, A.; Gross, B. J.; Day, L. A.; McDermott, A. E. J. Am.
- Chem. Soc. 2007, 129, 2338. (2) Chevelkov, V.; Faelber, K.; Schrey, A.; Rehbein, K.; Diehl, A.; Reif, B. J. Am. Chem. Soc. 2007, 129, 10195.
- (3) Pintacuda, G.; Giraud, N.; Pierattelli, R.; Bockmann, A.; Bertini, I.; Emsley, L. Angew. Chem., Int. Ed. Engl. 2007, 46, 1079.
- (4) Andronesi, O. C.; von Bergen, M.; Biernat, J.; Seidel, K.; Griesinger, C.; Mandelkow, E.; Baldus, M. J. Am. Chem. Soc. 2008, 130, 5922.

the I = 5/2 ¹⁷O nucleus, which has a low sensitivity due to its very low natural abundance (0.037%) and low resonance frequency ($\gamma = -3.628 \times 10^7$ rad T⁻¹ s⁻¹), as well as the presence of an electric quadrupole moment ($Q = -2.6 \times 10^{-30}$ m²). With isotopic enrichment, ¹⁷O NMR is being increasingly used in the study of inorganic materials,⁵ as well as organic and biological molecules.⁶ However, relatively few heteronuclear experiments involving oxygen have been presented. Cross polarization (CP) from ¹H or ²⁷Al to ¹⁷O has been demonstrated,⁷⁻¹¹ while ¹⁷O/¹H REDOR has been employed to investigate ¹⁷O-¹H proximities in zeolites HY and HZSM- $5.^{12}$ $^{13}C^{-17}O$ distance measurements via the SEDOR, REDOR, and REAPDOR techniques have also been reported for CO

- (5) Ashbrook, S. E.; Smith, M. E. Chem. Soc. Rev. 2006, 35, 718.
- (6) Wu, G. Prog. Nucl. Magn. Reson. Spectrosc. 2008, 52, 118.
- Walter, T. H.; Turner, G. L.; Oldfield, E. J. Magn. Reson. 1988, 76, (7)106.
- (8) Gann, S. L.; Baltisberger, J. H.; Wooten, E. W.; Zimmermann, H.; Pines, A. Bull. Magn. Reson. 1994, 16, 68.
- Kuroki, S.; Takahashi, A.; Ando, I.; Shoji, A.; Ozaki, T. J. Mol. Struct. 1994, 323, 197.
- (10) Ashbrook, S. E.; Wimperis, S. J. Magn. Reson. 2000, 147, 238.
- (11) Ashbrook, S. E.; Wimperis, S. Mol. Phys. 2000, 98, 1.
- (12) Peng, L.; Huo, H.; Liu, Y.; Grey, C. P. J. Am. Chem. Soc. 2007, 129, 335.

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chemisorbed on Pd metal,¹³ L-tyrosine,¹⁴ and asparagine monohydrate,¹⁵ as well as the differentiation of parallel and antiparallel β -sheets formed by L-alanyl-alanyl-alanine.¹⁶ Recently, experiments incorporating the symmetry-based recoupling of ${}^{1}\text{H}-{}^{17}\text{O}$ heteronuclear dipolar couplings have been presented, namely modified REDOR experiments that enable the measurement of intra- and intermolecular ¹H-¹⁷O distances in L-tyrosine \cdot HCl^{17,18} and ¹H \rightarrow ¹⁷O polarization transfer using the PRESTO-II sequence applied to glycine • HCl and Mg(OH)₂.¹⁹ Furthermore, the mobility of ¹⁷O nuclei and the assignment of 17 O and 51 V sites in anionic conductive materials (α -Bi₄V₂O₁₁ and γ -Bi₄V_{1.7}Ti_{0.3}O_{10.85}) using ¹⁷O/⁵¹V TRAPDOR experiments has been reported.²⁰ A few instances of two-dimensional (2D) heteronuclear experiments can also be found in the literature. In particular, 2D experiments devised to examine the relative orientation between the ¹⁷O quadrupole tensors and the O-H dipole tensors in glycine • HCl,¹⁹ Mg(OH)₂,^{19,21} and $Mg(OH)_x(OCH_3)_{2-x}^{21}$ have been reported. Recently, throughbond ²⁷Al/¹⁷O 2D HMQC correlation spectra for crystalline CA2(CaAl₂O₇) and vitreous CA(CaO-Al₂O₃) have also been presented.²² From the scarcity of ¹⁷O heteronuclear studies in the literature, it can be seen that considerable scope remains for further development of solid-state NMR experiments that probe proximities and connectivity with ¹⁷O nuclei, leading to additional constraints for structural elucidation.

Despite the importance of NH···O hydrogen bonds in organic and biological solids, to the best of our knowledge, ${}^{15}N{-}^{17}O$ solid-state NMR experiments have not previously been presented, with their proximate Larmor frequencies and combined low sensitivity representing difficult hurdles to overcome. In this paper, we demonstrate the feasibility of solid-state NMR experiments that utilize and probe ${}^{15}N{-}^{17}O$ dipolar and *J* couplings for two model compounds, glycine·²HCl and uracil.

2. Experimental and Computational Details

2.1. Materials and Synthesis. The sample of $[{}^{2}H(NH_{3}), 1{}^{13}C, {}^{15}N, {}^{17}O_{2}]glycine \cdot {}^{2}HCl was prepared as described previously {}^{23}$ by using commercially available $[1{}^{-13}C, {}^{15}N]glycine$ (Sigma-Aldrich, UK) and $[70\%{}^{-17}O]H_{2}O$ (CortecNet, France) as starting materials, with subsequent deuteration of the exchangeable protons (COOH, NH₃) being achieved by recrystallization from D₂O. $[1,3{}^{-15}N]Uracil$ (Sigma-Aldrich, UK) was enriched with $[70\%{}^{-17}O]H_{2}O$ according to the literature protocol, resulting in $[1,3{}^{-15}N,2,4{}^{-17}O]$ uracil.^{24,25}

2.2. Solid-State NMR. All experiments were performed on a Bruker AVANCE II+ NMR spectrometer, operating at ¹H, ¹³C, ¹⁵N, and ¹⁷O Larmor frequencies of 600, 151, 61, and 81 MHz, respectively, using a Bruker 3.2 mm triple-resonance magic-angle spinning (MAS) probe. Unless otherwise stated, ¹³C and ¹⁵N $\pi/2$ pulses of 5.0 μ s, corresponding to a nutation frequency (ν_1) equal to 50 kHz, were used, while "central-transition-selective" $\pi/2$ pulses of 5.0 μ s were used for ¹⁷O, corresponding to $\nu_1 = 16.7$ kHz (i.e., taking into account the scaling by (I + 1/2)⁻¹). SPINAL-64

- (13) Shore, S. E.; Ansermet, J. P.; Slichter, C. P.; Sinfelt, J. H. *Phys. Rev. Lett.* **1987**, *58*, 953.
- (14) Gullion, T. J. Magn. Reson. Ser. A 1995, 117, 326.
- (15) Chopin, L.; Vega, S.; Gullion, T. J. Am. Chem. Soc. 1998, 120, 4406.
- (16) Gullion, T.; Yamauchi, K.; Okonogi, M.; Asakura, T. *Macromolecules* 2007, 40, 1363.
- (17) Brinkmann, A.; Kentgens, A. P. M. J. Am. Chem. Soc. 2006, 128, 14758.
- (18) Brinkmann, A.; Kentgens, A. P. M. J. Phys. Chem. B 2006, 110, 16089.
- (19) van Beek, J. D.; Dupree, R.; Levitt, M. H. J. Magn. Reson. 2006, 179, 38.
- (20) Kim, N.; Grey, C. P. Science 2002, 297, 1317.
- (21) van Eck, E. R. H.; Smith, M. E. J. Chem. Phys. 1998, 108, 5904.

heteronuclear decoupling²⁶ was used with $\nu_1(^{1}\text{H}) = 100$ kHz. Ramped cross polarization²⁷ was employed for the acquisition of all ¹³C and ¹⁵N spectra with a contact time of 1.0 ms, a ¹H $\pi/2$ -pulse duration of 2.5 μ s, and (except where otherwise stated) a recycle delay of 4.0 s. For ¹⁷O experiments (except where otherwise stated), recycle delays of 0.5 s (glycine \cdot ²HCl) and 1.0 s (uracil) were used. RAPT²⁸ (n = 3, $\nu_{off} = 450$ kHz) was used for signal enhancement in ¹⁷O 1D experiments.

¹⁵N chemical shifts were referenced indirectly to neat liquid nitromethane (CH₃NO₂) by using powdered [¹⁵N]glycine as an external secondary reference ($\delta_{iso} = -347.4$ ppm). To convert to the chemical shift scale frequently used in protein NMR, where the reference is liquid ammonia at -50 °C, it is necessary to add 379.5 ppm to the given values. The ¹³C chemical shifts were referenced to tetramethylsilane (TMS, Si(CH₃)₄) by using powdered I-[1-¹³C]alanine as an external secondary reference ($\delta_{iso} = 177.9$ ppm). The ¹⁷O chemical shift scale was referenced to the ¹⁷O signal of H₂O set to 0.0 ppm.

2D ¹⁷O/¹⁵N R³-HMQC NMR spectra were acquired with the pulse sequence shown in Figure 1a, related versions of which have been used previously to record ¹H/¹⁴N, ¹H/²⁷Al, and ²³Na/³¹P correlation spectra.^{29,30} Å 16-step phase cycle was used, $\varphi_1 = +x, -x; \varphi_2 =$ +x, +x, +x, +x, +y, +y, +y, +y, -x, -x, -x, -x, -y, -y, -y, $-y; \varphi_3 = +y, +y, -y, -y; \varphi_4 = -y;$ receiver = -x, +x, +x, -x,+x, -x, -x, +x, -x, +x, -x, +x, -x, -x, +x, to select $\Delta p =$ ± 1 on the ¹⁷O $\pi/2$ pulse (φ_1), $\Delta p = \pm 2$ on the ¹⁷O π pulse (φ_2), and $\Delta p = \pm 1$ on the first ¹⁵N $\pi/2$ pulse (φ_3), where p is the coherence order. Sign discrimination in F_1 was achieved using the States hyper-complex processing method.³¹ A total of 512 (glycine^{•2}HCl) or 1024 (uracil) transients were coadded for each of 206 (glycine \cdot ²HCl) or 130 (uracil) t_1 slices, resulting in a total experimental time of 14.6 h (glycine^{•2}HCl) or 18.5 h (uracil). Spectral widths of 50.0 and 11.75 kHz (glycine · 2HCl) or 80.0 and 5.0 kHz (uracil) were employed along the F_2 and F_1 dimensions, respectively. The ¹⁵N carrier frequency was set at -299.1 (glycine • ²HCl) and -237.5 (uracil) ppm. R³ dipolar recoupling was achieved by applying pulses for $\tau/2 = 280\tau_r \approx 11.9$ ms at $\nu_1 = \nu_r$ = 23.5 kHz (glycine \cdot ²HCl) or $\tau/2 = 160\tau_r \approx 8.0$ ms at $\nu_1 = \nu_r =$ 20.0 kHz (uracil), where v_r and τ_r denote the MAS frequency and the rotor period, respectively.

¹⁷O/¹⁵N REDOR spectra of glycine ·²HCl were acquired at 22.5 kHz MAS, with a recycle delay of 2 s. The REDOR pulse sequence as employed previously for ²³Na/³¹P and ²⁷Al/³¹P REDOR experiments³² is shown in Figure 1b. The REDOR fraction ($\Delta S/S = (S - S_r)/S$) was determined from spectra acquired without (S_r) and with (S) the central ¹⁵N pulse, where the r subscript refers to the reduced signal intensity in the dephased (under the heteronuclear

- (22) Iuga, D.; Morais, C.; Gan, Z. H.; Neuville, D. R.; Cormier, L.; Massiot, D. J. Am. Chem. Soc. 2005, 127, 11540.
- (23) Wong, A.; Hung, I.; Howes, A. P.; Anupold, T.; Past, J.; Samoson, A.; Brown, S. P.; Smith, M. E.; Dupree, R. *Magn. Reson. Chem.* 2007, 45, S68.
- (24) Wu, G.; Dong, S.; Ida, R.; Reen, N. J. Am. Chem. Soc. 2002, 124, 1768.
- (25) Wu, G.; Dong, S. J. Am. Chem. Soc. 2001, 123, 9119.
- (26) Fung, B. M.; Khitrin, A. K.; Ermolaev, K. J. Magn. Reson. 2000, 142, 97.
- (27) Metz, G.; Wu, X. L.; Smith, S. O. J. Magn. Reson. Ser. A 1994, 110, 219.
- (28) Kwak, H. T.; Prasad, S.; Clark, T.; Grandinetti, P. J. J. Magn. Reson. 2003, 160, 107.
- (29) Gan, Z. H.; Amoureux, J. P.; Trebosc, J. Chem. Phys. Lett. 2007, 435, 163.
- (30) Trebosc, J.; Hu, B.; Amoureux, J. P.; Gan, Z. J. Magn. Reson. 2007, 186, 220.
- (31) States, D. J.; Haberkorn, R. A.; Ruben, D. J. J. Magn. Reson. 1982, 48, 286.
- (32) Chan, J. C. C.; Eckert, H. J. Magn. Reson. 2000, 147, 170.
- (33) Gullion, T. Chem. Phys. Lett. 1995, 246, 325.
- (34) Goldbourt, A.; Vega, S.; Gullion, T.; Vega, A. J. J. Am. Chem. Soc. 2003, 125, 11194.



Figure 1. Pulse sequences for the (a) \mathbb{R}^3 -HMQC, (b) REDOR, (c) CP/ REAPDOR, and heteronuclear spin—echo experiments with (d) one pulse and (e) CP excitation of the observed nucleus. The coherence transfer pathway diagram for (a) is shown below the pulse sequence. For the (b) REDOR and (c) CP/REAPDOR experiments, the reduced-intensity dephased (S_r) and reference (S) spectra correspond to experiments (b) without (S_r) and with (S) the central ¹⁵N pulse and (c) with (S_r) and without (S) the ¹⁷O pulse.

dipolar coupling) spectrum. A two-step phase cycle was used, $\varphi_1 = +x, -x$; receiver = +x, -x. The ¹⁵N and ¹⁷O π pulses were of duration 8 μ s, while the initial ¹⁷O direct excitation pulse was of duration 4 μ s. A total of 1152 transients were coadded for the S_r and S experiments recorded for each τ duration.

¹⁵N/¹⁷O CP/REAPDOR spectra were acquired at various MAS frequencies observing on the ¹⁵N channel and using pulses of duration one-third of a rotor period, $\tau_r/3$, on the ¹⁷O channel with $\nu_1(^{17}O)$ approximately equal to 75 kHz. The REAPDOR fraction ($\Delta S/S = (S - S_r)/S$) was determined from spectra acquired with (S_r) and without (S) the ¹⁷O adiabatic pulse. The REAPDOR pulse sequence, as previously used for X/Y experiments,^{14–16,33–35} is

shown in Figure 1c and employs an eight-step phase cycle, $\varphi_1 = +y, -y; \varphi_2 = +x, +x, -x, -x, +y, +y, -y, -y;$ receiver = +x, -x, -x, +x, +y, -y, -y, +y, that selects $\Delta p = \pm 1$ on the ¹H $\pi/2$ pulse (φ_1) and $\Delta p = -1$ on the ¹⁵N pulses (φ_2). Eight transients were coadded for the S_r and S experiments recorded for each τ duration. In both REDOR and REAPDOR experiments, the S_r and S spectra were recorded consecutively for each evolution interval, and the rotor-synchronized π -pulses on ¹⁵N followed the XY-8 (+x, +y, +x, +x, +y, +x, +y, +x, +y, +x, +y, +x, +y, +x, +y, +x, +x, +x, +x, +x, +y, +x, +x

A ¹⁷O/¹³C and a ¹⁷O/¹⁵N heteronuclear spin—echo experiment was performed on glycine ·²HCl and uracil, respectively, using the pulse sequence shown in Figure 1d; such a pulse sequence has been used previously to determine ²⁷Al–³¹P^{37,38} and ⁷¹Ga–³¹P³⁹ J couplings. ¹⁷O homonuclear spin—echo experiments were performed by omitting the π pulse on the nonobserved (i.e., ¹³C or ¹⁵N) channel. An eight-step phase cycle was used, $\varphi_1 = -x, +x; \varphi_2 =$ +y, +y, -x, -x, -y, -y, +x, +x; receiver = +x, -x, -x, +x, +x, -x, -x, +x, to select $\Delta p = \pm 1$ and ± 2 on the ¹⁷O $\pi/2$ (φ_1) and π pulses (φ_2), respectively. In total, 128 (6 s recycle delay) and 1152 (7.5 s recycle delay) transients were coadded for each τ increment for glycine ·²HCl and uracil, respectively.

A ¹³C/¹⁷O and a ¹⁵N/¹⁷O heteronuclear spin—echo experiment was performed on glycine ²HCl and uracil, respectively, using the pulse sequence depicted in Figure 1e employing the same phase cycling as described above for the ¹⁷O/¹³C or ¹⁷O/¹⁵N experiment in Figure 1d. ¹³C or ¹⁵N homonuclear spin—echo experiments were performed by omitting the π pulse on the nonobserve (i.e., ¹⁷O) channel. In total, 32 (6 s recycle delay) and 256 (7.5 s recycle delay) transients were coadded for each τ increment for the ¹³C/¹⁷O glycine ²HCl and ¹⁵N/¹⁷O uracil experiments, respectively.

Where data corresponding to the quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$ are presented, the homonuclear and heteronuclear spin—echo spectra were recorded consecutively for each evolution interval. $S_{\text{HET}}(\tau)$ and $S_{\text{HOM}}(\tau)$ integrals were taken over the respective resolved peaks after Fourier transformation with respect to t_2 . Errors on fitted parameters were determined using the covariance method as described in ref 40.

2.3. Computational Details. First-principles calculations were performed using the CASTEP⁴¹ software package, which implements density functional theory using a plane-wave basis set and the pseudopotential approach and is thus applicable to periodic systems.

Starting with the crystal structures of glycine \cdot HCl⁴² and uracil⁴³ (CSD reference codes GLYHCL01 and URACIL), geometry optimizations (at fixed lattice parameters) were performed. For glycine \cdot HCl, the Trouiller–Martins norm-conserving pseudopotentials⁴⁴ with a plane-wave cutoff at 1200 eV and a *k*-point density of 0.07 Å⁻¹ were used. For a first optimization where only the hydrogen atoms were relaxed, high forces (up to 4.2 eV/Å) remained, such that a full relaxation of all atoms was performed. The geometry optimization of the uracil crystal structure has been described in ref 45, where all the atoms in the unit cell were relaxed.

- (36) Gullion, T.; Baker, D. B.; Conradi, M. S. J. Magn. Reson. 1990, 89, 479.
- (37) Amoureux, J. P.; Trebosc, J.; Wiench, J. W.; Massiot, D.; Pruski, M. Solid State Nucl. Magn. Reson. 2005, 27, 228.
- (38) Massiot, D.; Fayon, F.; Alonso, B.; Trebosc, J.; Amoureux, J. P. J. Magn. Reson. 2003, 164, 160.
- (39) Montouillout, V.; Morais, C. M.; Douy, A.; Fayon, F.; Massiot, D. Magn. Reson. Chem. 2006, 44, 770.
- (40) Pham, T. N.; Griffin, J. M.; Masiero, S.; Lena, S.; Gottarelli, G.; Hodgkinson, P.; Fillip, C.; Brown, S. P. *Phys. Chem. Chem. Phys.* 2007, 9, 3416.
- (41) Clark, S. J.; Segall, M. D.; Pickard, C. J.; Hasnip, P. J.; Probert, M. J.; Refson, K.; Payne, M. C. Z. Kristallogr. 2005, 220, 567.
- (42) Al-Karaghouli, A. R.; Cole, F. E.; Lehmann, M. S.; Miskell, C. F.; Verbist, J. J.; Koetzle, T. F. J. Chem. Phys. 1975, 63, 1360.
- (43) Stewart, R. F.; Jensen, L. H. Acta Crystallogr. 1967, 23, 1102.
- (44) Troullier, N.; Martins, J. L. Phys. Rev. B 1991, 43, 1993.
- (45) Uldry, A. C.; et al. J. Am. Chem. Soc. 2008, 130, 945.

⁽³⁵⁾ Gullion, T.; Vega, A. J. Prog. Nucl. Magn. Reson. Spectrosc. 2005, 47, 123.

J couplings were calculated as described by Joyce et al. using a developer version of the CASTEP software package:⁴⁶ Trouiller-Martins norm-conserving pseudopotentials⁴⁴ are used together with the projector augmented wave (PAW)⁴⁷ technique in order to recover the all-electron accuracy. The exchange-correlation functional employed was the Perdew-Burke-Ernzerhof (PBE)⁴⁸ implementation of the generalized gradient approximation. The Jcoupling calculations were performed with a plane-wave cutoff of 1100 eV and a k-point sampling grid density of 0.09 (glycine HCl) or 0.08 (uracil) $Å^{-1}$. The J couplings are computed by considering one nucleus as a perturbation, and it is therefore necessary to multiply the size of the original crystal unit cell until the values of the couplings are converged. A $2 \times 2 \times 1$ supercell (192 atoms) and a $2 \times 2 \times 2$ supercell (384 atoms) were found to be sufficient for the J couplings to be well converged within ± 0.1 Hz for glycine · HCl and uracil, respectively. For uracil, the calculation time for the J couplings varied between 27 and 41 h on 24 nodes of the AMD Opteron cluster at St. Andrews University, depending on the perturbing nucleus.

3. Molecular Structures and 1D MAS NMR Spectra

The arrangement of glycine and uracil molecules in the geometrically optimized (CASTEP) crystal structures of glycine \cdot HCl⁴² and uracil⁴³ is illustrated in Figure 2. The intramolecular C–O distances in glycine \cdot HCl, as well as the closest intra- and intermolecular N–O distances in glycine \cdot HCl and uracil, are indicated. The N–O distances are listed in Table 1, which also specifies the corresponding dipolar coupling constants in hertz:

$$D_{jk} = -\frac{\mu_0 \gamma_j \gamma_k \hbar}{8\pi^2 r_{ik}^3} \tag{1}$$

where r_{jk} is the internuclear distance between spins *j* and *k*, and γ_j and γ_k are the respective gyromagnetic ratios of *j* and *k*.



Figure 2. Representations of the geometrically optimized (CASTEP) crystal structures of (a) glycine •HCl⁴² and (b) uracil.⁴³ Color coding: H, white; C, gray; N, blue; O, red; Cl, green. The closest intramolecular (in black) and intermolecular (in red) distances are indicated. The isotopically enriched sites in both molecules are labeled within the insets.

Table 1. Intra- and Intermolecular^a N-O Proximities^b in Glycine HCl and Uracil

sample	j	k	r _{jk} /Å	<i>ID_{jk}I ∕</i> Hz
glycine • HCl	Ν	01	2.74	80
	Ν	01	2.94	65
	Ν	O2	3.65	34
uracil	N1	O2	2.30	135
	N1	04	2.83	73
	N3	O2	2.30	136
	N3	O4	2.28	139
	N3	04	2.84	72

^{*a*} Intermolecular distances are highlighted using boldface type. ^{*b*} Distances are from the geometrically optimized (CASTEP) crystal structures.

For the nitrogen atom in glycine ·HCl, the closest intermolecular N–O proximity of 2.94 Å to O1 (C=O) is longer than the intramolecular N–O proximity of 2.74 Å. For O2 (C–OH), the closest proximity is the intramolecular N-O distance of 3.65 Å, with the nearest intermolecular proximity being at 4.15 Å. In uracil, there are three short intramolecular N–O distances (2.28-2.30 Å), with N3 being sandwiched between two C=O groups and hence having two close intramolecular N-O proximities as compared to the one for N1. Both N1 and N3 form intermolecular NH····O hydrogen bonds to O4 with N-O distances of 2.83 and 2.84 Å (Figure 2b). While O4 takes part in NH····O hydrogen bonds with both N1 and N3, O2 forms weak CH···O hydrogen bonds, which have been the subject of a recent combined experimental and first-principles computational study;⁴⁵ note the different atom numbering convention in ref 45 as compared to the present work.

It is evident that the N–O proximities in glycine \cdot HCl and uracil do not correspond to well-isolated spin pairs. In this case, it is informative to consider the root-sum-squared dipolar coupling:⁴⁹

$$D_{\text{rss},j} = \sqrt{\sum_{k \neq j} D_{jk}^2}$$
(2)

which, in analogy to a second moment analysis, is a measure of the "effective coupling" at a particular site. Specifically, $D_{rss,j}$ is related to the second moment (for heteronuclear dipolar coupled nuclei) by⁵⁰

$$M_2 = (4/15)I(I+1)(2\pi)^2 D_{\rm rss\,i}^2 \tag{3}$$

where *I* is the spin quantum number of the nuclear spins coupled to nucleus *j*. $D_{rss,j}$ values for the dipolar couplings of the ¹⁵N nuclei in glycine•HCl and uracil to the surrounding oxygen nuclei (considering N–O distances up to 6.0 Å) are listed in Table 2. However, it is to be noted that the oxygen sites are only partially ¹⁷O labeled. Assuming the isotopic labels are randomly distributed, Zorin et al. have shown (see eq 6 of ref 51) that

$$D_{\text{rss},i}(\rho) = \sqrt{\rho} D_{\text{rss},i} \tag{4}$$

where ρ denotes the proportion of labeled nuclei.

- (46) Joyce, S. A.; Yates, J. R.; Pickard, C. J.; Mauri, F. J. Chem. Phys. **2007**, *127*, 204107.
- (47) Blochl, P. E. Phys. Rev. B 1994, 50, 17953.
- (48) Perdew, J. P.; Burke, K.; Ernzerhof, M. Phys. Rev. Lett. **1996**, 77, 3865.
- (49) Zorin, V. E.; Brown, S. P.; Hodgkinson, P. Mol. Phys. 2006, 104, 293.
- (50) van Wullen, L.; Muller, U.; Jansen, M. Chem. Mater. 2000, 12, 2347.

Table 2. N–O Root-Sum-Squared Dipolar Couplings Determined from the Geometrically Optimized (CASTEP) Crystal Structures of Glycine • HCl and Uracil

sample	j	k	$D_{\rm rss,}/{\rm Hz}^a$
glycine · HCl	Ν	01	108
	Ν	O2	58
	Ν	01, 02	123
	O1	Ν	108
	O2	Ν	58
uracil	N1	O2	149
	N1	O4	87
	N1	O2, O4	173
	N3	O2	142
	N3	O4	166
	N3	02, 04	219



Figure 3. ¹⁷O MAS NMR spectra of (a) [²H(NH₃),1-¹³C,¹⁵N,¹⁷O₂]glycine·²HCl ($\nu_r = 22.5$ kHz) and (b) [¹⁵N₂,¹⁷O₂]uracil ($\nu_r = 20.0$ kHz). (c) ¹⁵N CP/MAS spectrum of [¹⁵N₂,¹⁷O₂]uracil at $\nu_r = 10.0$ kHz. All spectra were recorded at 14.1 T (ν_0 (¹H) = 600 MHz). A total of (a) 256, (b) 2400, and (c) 4 transients were coadded. RAPT²⁸ (n = 3, $\nu_{off} = 450$ kHz) was used for signal enhancement of ¹⁷O spectra.

¹³C and ¹⁵N CP/MAS spectra (not shown) of the labeled glycine \cdot^{2} HCl sample both show a single resonance at 172.6 and -343.5 ppm, respectively, while two well-resolved quadrupolar patterns corresponding to the two distinct COO²H group ¹⁷O sites are observed in a ¹⁷O MAS spectrum at 14.1 T (Figure 3a). The ¹⁷O resonances centered at approximately 280 and 130 ppm are assigned to the C=O (O1) and C-O²H (O2)

Table 3. ¹⁷O Isotropic Chemical Shift and Quadrupolar Parameters for Glycine ^{•2}HCI and Uracil

sample	site	$\delta_{\rm iso}$ /ppm	<i>C₀</i> /MHz ^c	η_Q
glycine • 2HCla	01	334	8.34	0.00
	O2	175	7.48	0.24
uracil ^b	O2	240	7.62	0.50
	O4	275	7.85	0.55

^{*a*} Parameters taken from ref 23. ^{*b*} Parameters taken from ref 25. ^{*c*} C_Q = $e^2 q Q/h$, where eq is the electric field gradient and Q is the nuclear quadrupole moment.

sites, respectively.^{23,52} In contrast, for the labeled uracil sample, the signals from the O2 and O4 sites cannot be resolved in a ¹⁷O MAS NMR spectrum at 14.1 T, which shows a single broadened pattern centered at 230 ppm (Figure 3b). Overlap of the signals is caused by the similarity in chemical shifts and quadrupolar parameters for the two oxygen nuclei, which have previously been studied by ¹⁷O MQMAS.²⁵ The isotropic chemical shifts and quadrupolar parameters for the ¹⁷O sites in glycine ^{•2}HCl and uracil are listed in Table 3. The ¹⁵N CP/MAS spectrum of uracil shows two signals at -244.2 and -222.2 ppm which are assigned to the N1 and N3 sites (Figure 3c).⁴⁵

4. Two-Dimensional Through-Space Correlation Experiments

Recently, heteronuclear multiple-quantum correlation (HM-QC)⁵³ solid-state NMR experiments have been presented that correlate the chemical shift evolution of coupled spin I = 1/2and quadrupolar nuclei, with applications to ¹H/¹⁴N,^{29,54} ¹³C/¹⁴N, ^{55,56} ³¹P/²³Na, ³⁰ ¹H/²⁷Al, ³⁰ and ³¹P/²⁷Al³⁰ having been reported. One demonstrated method for establishing such heteronuclear correlations is the use of rotary-resonance recoupling $(\mathbb{R}^3)^{57}$ to recouple the heteronuclear dipolar coupling that would otherwise be averaged to zero by MAS. In the R³ method, radio frequency (rf) irradiation of duration $N\tau_r$ reintroduces homonuclear dipolar coupling, heteronuclear dipolar coupling, and/or chemical-shift anisotropy depending on the ratio of the applied rf frequency and the rotation frequency, $v_1 = nv_r$ (n =1/2, 1, 2).²⁹ The $n = 1 R^3$ condition is most efficient and recouples both the homo- and heteronuclear dipolar interactions; hence, it is employed in the present cases since the effect of homonuclear ¹⁵N couplings is generally expected to be weak. The application of R^3 has been thoroughly discussed elsewhere, ^{29,30,57} and its comparison with other heteronuclear dipolar recoupling methods has recently been reported.58 The simplicity of applying R³ stems from the limited number of variables required for its optimization. Having chosen which $v_1 = nv_r R^3$ condition to employ, only the recoupling pulse length $(\tau/2)$ remains to be determined. In both instances presented herein, the maximum ¹⁷O R³-HMQC intensity (\sim 5–7% compared to a spin-echo experiment) was observed when $\tau/2$ was ap-

- (53) Muller, L. J. Am. Chem. Soc. 1979, 101, 4481.
- (54) Cavadini, S.; Abraham, A.; Bodenhausen, G. Chem. Phys. Lett. 2007, 445, 1.
- (55) Gan, Z. H. J. Magn. Reson. 2007, 184, 39.
- (56) Cavadini, S.; Antonijevic, S.; Lupulescu, A.; Bodenhausen, G. ChemPhysChem 2007, 8, 1363.
- (57) Oas, T. G.; Griffin, R. G.; Levitt, M. H. J. Chem. Phys. 1988, 89, 692.
- (58) Hu, B.; Trebosc, J.; Amoureux, J. P. J. Magn. Reson. 2008, 192, 112.

⁽⁵¹⁾ Zorin, V. E.; Brown, S. P.; Hodgkinson, P. J. Chem. Phys. 2006, 125, 144508.

⁽⁵²⁾ Pike, K. J.; Lemaitre, V.; Kukol, A.; Anupold, T.; Samoson, A.; Howes, A. P.; Watts, A.; Smith, M. E.; Dupree, R. J. Phys. Chem. B 2004, 108, 9256.



Figure 4. 2D ¹⁷O/¹⁵N R³-HMQC NMR spectra of (a) [²H(NH₃),1-¹³C,¹⁵N, ¹⁷O₂]glycine·²HCl and (b) [¹⁵N₂,¹⁷O₂]uracil with skyline projections along F_1 and F_2 . The base contours are set at 12% and 20% of the maximum intensity for spectra a and b, respectively. The ¹⁷O MAS spectrum (dotted red line) of [²H(NH₃), 1-¹³C,¹⁵N,¹⁷O₂]glycine·²HCl is superimposed over the F_2 projection in spectrum a for comparison.

proximately equal to the inverse of the effective dipolar coupling (see section 3).

A 2D ¹⁷O/¹⁵N R³-HMQC NMR spectrum for glycine • ²HCl acquired using the sequence in Figure 1a is shown in Figure 4a. A clear correlation between the ¹⁵N site and both ¹⁷O resonances can be observed, attesting to the proximities between the nitrogen and both oxygen nuclei. The projection along the ¹⁷O dimension provides an accurate representation of the O1 pattern when compared to the ¹⁷O MAS spectrum, whereas the O2 signal shows less resemblance-this may be due to the poorer signal intensity. The relative intensities of the two correlation peaks show a qualitative correspondence with the relative distance between the ¹⁵N and the O1 (2.7 Å) and O2 (3.7 Å) sites (Figure 2a) as well as the effective dipolar couplings considering all N-O1 and N-O2 distances under 6 Å, $D_{rss,j}(N-O1) = 108$ and $D_{rss,j}(N-O2) = 58$ Hz (see Table 2). The ¹⁷O/¹⁵N R³-HMQC spectrum for uracil also shows two distinct correlation peaks (Figure 4b). However, in this instance the two oxygen sites cannot be distinguished (see the ¹⁷O MAS spectrum in Figure 3b); hence, it can only be stated that there are spatial proximities between each N site and at least one O site. In order to separate the different quadrupolar patterns, a high-resolution method such as MQMAS⁵⁹ or STMAS⁶⁰ would have to be included into the HMQC experiment, as has been suggested in the literature.⁶¹ The spectra presented here demonstrate the feasibility of observing ¹⁵N-¹⁷O R³-HMQC heteronuclear correlations for N–O spatial proximities out to 3.7 Å; i.e., heteronuclear correlations between ¹⁵N and ¹⁷O can be obtained with dipolar couplings as small as ~35 Hz.

5. Quantitative Measurement of Root-Sum-Squared Through-Space Dipolar Couplings

The above section has shown that heteronuclear spatial proximities can be observed in a semiquantitative fashion via 2D correlation experiments. This section presents ${}^{17}O{-}^{15}N$ rotational-echo double-resonance (REDOR)^{62,63} and ${}^{15}N{-}^{17}O$ rotational-echo adiabatic-passage double-resonance (REA-PDOR)^{14,33,35} experimental buildup data for glycine ${}^{2}HCl$ and uracil, from which root-sum-squared dipolar couplings can be quantitatively determined.

5.1. ¹⁷O-¹⁵N REDOR Experiments. The REDOR fraction is given as $\Delta S/S = (S - S_r)/S$, where the r subscript in S_r refers to the reduced signal intensity in the dephased (under the heteronuclear dipolar coupling) spectrum, while S refers to the reference spectrum. For a pair of dipolar-coupled spin I = 1/2nuclei, it is well established that the REDOR fraction depends only on the evolution time ($\tau = N\tau_r$) and the dipolar coupling constant D_{ik} , with this simple relation explaining the very widespread application of the REDOR experiment.⁶³ However, the ¹⁵N and ¹⁷O nuclei in glycine ·HCl and uracil do not exist as well-isolated spin pairs (see section 3). For IS_n systems, the analysis is considerably complicated by the dependence of the REDOR response on the relative orientation of the internuclear vectors corresponding to the different IS heteronuclear dipolar couplings.^{64–68} However, starting with the expression of Mueller for the initial REDOR buildup (for $\Delta S/S < 0.2$, where $\cos(x)$ can be expressed as $1 - x^2/2$,⁶⁹

$$\Delta S/S = \frac{16N^2\tau_{\rm r}^2}{15} \sum_{k\neq i} D_{jk}^2 = \frac{16N^2\tau_{\rm r}^2}{15} D_{{\rm rss},i}^2 \tag{5}$$

Eckert and co-workers have shown that the initial buildup of a REDOR experiment ($\Delta S/S < 0.2$) can be fit to the following parabolic function (using the definition of the second moment in eq 3):^{70–72}

$$\Delta S/S = \frac{N^2 \tau_{\rm r}^2}{\pi^2 I(I+1)} M_2 \tag{6}$$

- (59) Frydman, L.; Harwood, J. S. J. Am. Chem. Soc. 1995, 117, 5367.
- (60) Gan, Z. H. J. Am. Chem. Soc. 2000, 122, 3242.
- (61) Amoureux, J. P.; Trebosc, J.; Wiench, J.; Pruski, M. J. Magn. Reson. 2007, 184, 1.
- (62) Gullion, T.; Schaefer, J. J. Magn. Reson. 1989, 81, 196.
- (63) Gullion, T. Concepts Magn. Reson. 1998, 10, 277.
- (64) Naito, A.; Nishimura, K.; Tuzi, S.; Saito, H. Chem. Phys. Lett. 1994, 229, 506.
- (65) Goetz, J. M.; Schaefer, J. J. Magn. Reson. 1997, 127, 147.
- (66) Brouwer, E. B.; Gougeon, R. D. M.; Hirschinger, J.; Udachin, K. A.; Harris, R. K.; Ripmeester, J. A. Phys. Chem. Chem. Phys. 1999, 1, 4043.
- (67) Nishimura, K.; Naito, A.; Tuzi, S.; Saito, H. J. Phys. Chem. B 1999, 103, 8398.
- (68) Fyfe, C. A.; Lewis, A. R. J. Phys. Chem. B 2000, 104, 48.
- (69) Mueller, K. T. J. Magn. Reson. Ser. A 1995, 113, 81.
- (70) Bertmer, M.; Eckert, H. Solid State Nucl. Magn. Reson. 1999, 15, 139.
- (71) Chan, J. C. C.; Bertmer, M.; Eckert, H. J. Am. Chem. Soc. 1999, 121, 5238.
- (72) Bertmer, M.; Zuchner, L.; Chan, J. C. C.; Eckert, H. J. Phys. Chem. B 2000, 104, 6541.



Figure 5. Experimental ¹⁷O/¹⁵N REDOR curves, $\Delta S/S = (S - S_r)/S$, as a function of the dephasing time for the O1 (C=O) (filled squares) and O2 (O²H) (open squares) resonances of [²H(NH₃),1-¹³C,¹⁵N,¹⁷O₂]glycine+²HCl. The best fits to eq 5 for $\Delta S/S \le 0.2$ (horizontal dashed line) are shown as solid lines.

where *I* is the spin-quantum number of the nonobserve nucleus. The validity of eq 6 has been demonstrated by simulations of IS_n systems with different geometries^{70,71} and ²³Na/³¹P and ²⁷Al/³¹P REDOR experiments on sodium and aluminum phosphates for which the fitted second moments are within 10% of those calculated using the known crystal structures.³² This approach has subsequently been employed in the quantitative analysis of ²⁹Si/¹H, ²³Na/³¹P, and ⁶Li/³¹P REDOR data for zeolites,⁷³ glasses,^{74,75} and lithium ion site dynamics.⁷⁶ Moreover, Strojek et al. have shown that the approach can be extended to REDOR experiments for a I = 3/2 nonobserve nucleus, whereby it is necessary to include a coefficient (determined from simulations that require only an estimate of the C_Q and η_Q of the quadrupolar nucleus) to take into account the incomplete inversion of the satellite transitions.⁷⁷

Experimental ¹⁷O/¹⁵N REDOR buildup data obtained using the pulse sequence in Figure 1b for the carbonyl ¹⁷O (O1) and hydroxyl ¹⁷O (O2) resonances of glycine ·²HCl are shown in Figure 5. The slower dephasing for the hydroxyl resonance consistent with the $D_{rss,j}$ values in Table 2—is evident when comparing the buildup curves for O1 and O2 in Figure 5. Best fits ($\Delta S/S < 0.2$) to eq 5—shown as solid lines—yield fitted values for $D_{rss,j}$ of 89 ± 9 and 46 ± 6 Hz for the carbonyl and hydroxyl resonances, respectively, corresponding to 82% and 79% of the values determined from the crystal structure (see Table 2).

5.2. ¹⁵N $^{-17}$ **O REAPDOR Experiments.** The REAPDOR experiment (Figure 1c) is a modification of the REDOR experiment that allows the quantitative determination of the heteronuclear dipolar coupling between spin I = 1/2 nuclei (observe) and quadrupolar nuclei. Considering half-integer quadrupolar nuclei, REAPDOR experiments have been performed for various combinations of nuclei: ¹H/²³Na,⁷⁸ ¹H/²⁷Al,⁷⁸⁻⁸¹ ¹³C/¹¹B,⁸²

- (73) Magusin, P. C. M. M.; Zorin, V. E.; Aerts, A.; Houssin, C. J. Y.; Yakovlev, A. L.; Kirschhock, C. E. A.; Martens, J. A.; van Santen, R. A. J. Phys. Chem. B 2005, 109, 22767.
- (74) Santagneli, S. H.; de Araujo, C. C.; Strojek, W.; Eckert, H.; Poirier, G.; Ribeiro, S. J. L.; Messaddeq, Y. J. Phys. Chem. B 2007, 111, 10109.
- (75) Strojek, W.; Fehse, C. M.; Eckert, H.; Ewald, B.; Kniep, R. Solid State Nucl. Magn. Reson. 2007, 32, 89.
- (76) Cahill, L. S.; Kirby, C. W.; Goward, G. R. J. Phys. Chem. C 2008, 112, 2215.
- (77) Strojek, W.; Kalwei, M.; Eckert, H. J. Phys. Chem. B 2004, 108, 7061.
- (78) Holland, G. P.; Alam, T. M. Phys. Chem. Chem. Phys. 2005, 7, 1739.

 ${}^{13}C/{}^{17}O, {}^{14-16,34} {}^{13}C/{}^{27}Al, {}^{79} {}^{13}C/{}^{67}Zn, {}^{34} {}^{15}N/{}^{23}Na, {}^{78} {}^{15}N/{}^{27}Al, {}^{78} {}^{29}Si/{}^{11}B, {}^{50,82,83}{}^{29}Si/{}^{23}Na, {}^{84}{}^{29}Si/{}^{27}Al, {}^{85}{}^{31}P/{}^{27}Al, {}^{86-8831}P/{}^{45}Sc, {}^{8931}P/{}^{51}V, {}^{90}$ and ${}^{31}P/{}^{59}Co. {}^{91}$ Their applications include a tripeptide, 16 a metal-nucleotide complex, 91 zeolites, ${}^{78-81,84,85}$ ceramics, 50,82 glasses, ${}^{83,87-89}$ and vanadium-substituted polyoxoanionic solids. 90

In the REDOR experiment, dephasing due to the heteronuclear dipolar coupling relies on π inversion pulses applied every half a rotor period, so as to recouple the dipolar coupling that would otherwise be averaged to zero by MAS over one complete rotor period.^{62,63} For quadrupolar nuclei, it is often the case that the quadrupolar broadening extends over megahertz for powdered samples, such that at usual rf nutation frequencies it is not possible to achieve a uniform inversion of the broad distribution of orientation-dependent resonances. In the REAP-DOR experiment, an enhanced dephasing as compared to a REDOR experiment^{15,50} is achieved by replacing the π pulse on the quadrupolar nucleus by a so-called adiabatic-passage pulse. Specifically, such a pulse achieves a transfer of populations among the m states of the quadrupolar nuclei by means of the avoided level crossings that link different eigenstates of the quadrupolar Hamiltonian. These avoided level crossings can occur when the quadrupolar splitting changes sign, i.e., goes through zero, as a consequence of MAS-the $\cos(\omega_r t)$ and $\cos(2\omega_r t)$ dependence means that there are two or four zerocrossings per rotor period, depending on the orientation of the quadrupolar tensor.^{35,92} When a spin changes its eigenstate at such a zero-crossing, the passage is said to be adiabatic, with the likelihood of an adiabatic passage depending on the adiabaticity parameter:93-95

$$\alpha = \nu_1^2 / \nu_0 \nu_r \tag{7}$$

where

$$v_Q = 3C_Q / [2I(2I-1)]$$

= $3C_Q / 20$ for $I = 5/2$ (8)

For a spin I = 5/2 nucleus, experiment and simulation has shown that the adiabaticity parameter must exceed 0.55.^{15,34} The adiabatic pulse is set to be of duration $\tau_t/3$, since this corresponds to the maximizing of the number of crystallites in a powder making a single zero-crossing (NB: an even number of crossings)

- (79) van Wullen, L.; Koller, H.; Kalwei, M. Phys. Chem. Chem. Phys. **2002**, *4*, 1665.
- (80) Ganapathy, S.; Kumar, R.; Delevoye, L.; Amoureux, J. P. *Chem. Commun.* **2003**, 2076.
- (81) Kalwei, M.; Koller, H. Solid State Nucl. Magn. Reson. 2002, 21, 145.
- (82) van Wullen, L.; Jansen, M. J. Mater. Chem. 2001, 11, 223.
- (83) van Wullen, L.; Schwering, G. Solid State Nucl. Magn. Reson. 2002, 21, 134.
- (84) Ganapathy, S.; Vega, S. J. Am. Chem. Soc. 1998, 120, 1078.
- (85) Ganapathy, S.; Kumar, R.; Montouillout, V.; Fernandez, C.; Amoureux, J. P. Chem. Phys. Lett. 2004, 390, 79.
- (86) van Wullen, L.; Tricot, G.; Wegner, S. Solid State Nucl. Magn. Reson. 2007, 32, 44.
- (87) Zhang, L.; Eckert, H. J. Phys. Chem. B 2006, 110, 8946.
- (88) Wegner, S.; van Wullen, L.; Tricot, G. J. Non-Cryst. Solids 2008, 354, 1703.
- (89) Mohr, D.; de Camargo, A. S. S.; de Araujo, C. C.; Eckert, H. J. Mater. Chem. 2007, 17, 3733.
- (90) Huang, W. L.; Vega, A. J.; Gullion, T.; Polenova, T. J. Am. Chem. Soc. 2007, 129, 13027.
- (91) Grant, C. V.; Frydman, V.; Harwood, J. S.; Frydman, L. J. Am. Chem. Soc. 2002, 124, 4458.
- (92) Ba, Y.; Kao, H. M.; Grey, G. P.; Chopin, L.; Gullion, T. J. Magn. Reson. 1998, 133, 104.

during a rotor period would leave the spin in the same eigenstate). $^{92}\,$

As compared to the REDOR experiment,⁶³ the REAPDOR fraction, $\Delta S/S = (S - S_r)/S$, is not a simple function of the dipolar coupling constant because the quadrupolar spin flips are more complex than a simple inversion of their *z*-component, with the adiabatic pulse inducing partial transitions among the magnetic spin states *m* of the quadrupolar nuclei, rather than effecting a uniform 180° inversion.⁹⁶ However, an advantage of the REAPDOR experiment is that there is only a slight dependence of the REAPDOR fraction on the orientation of the quadrupolar tensor with respect to the internuclear vector, with the latter being difficult to determine experimentally.⁸¹ Indeed, Goldbourt et al. have shown that for a spin I = 1/2 nucleus coupled to a spin I = 5/2 nucleus, there exists a universal REAPDOR curve (valid when $\alpha > 0.55$ for an adiabatic pulse of duration $\tau_r/3$):³⁴

$$\Delta S/S = 0.63(1 - \exp[-(3.0D_{jk}\tau)^2]) + 0.2(1 - \exp[-(0.7D_{jk}\tau)^2]) \quad (9)$$

Specifically, simulated curves for C_Q between 1 and 13 MHz, η_Q between 0 and 1, ν_1 between 40 and 150 kHz, and ν_r between 3 and 20 kHz (for a range of orientations of the quadrupolar tensor with respect to the internuclear vector) were shown to not deviate (for $D_{jk}\tau < 0.5$) more than 15% from the universal curve.³⁴

Experimental ¹⁵N/¹⁷O CP/REAPDOR buildup data for glycine²HCl and uracil are shown in Figure 6. Experiments were performed at different MAS frequencies, with the adiabaticity parameter in all cases exceeding 0.55: for glycine • ²HCl, $v_r = 3.5 \ (\alpha \approx 1.3), 5.0 \ (\alpha \approx 0.9), and 8.0 \ (\alpha \approx 0.6) \text{ kHz; for}$ uracil, $\nu_r = 6.5$ ($\alpha \approx 0.75$) and 7.0 ($\alpha \approx 0.70$) kHz; see Table 3 for C_0 values, $\nu_1({}^{17}\text{O}) = 75$ kHz. Best fits to eq 9 are shown as solid lines, with dashed lines corresponding to a $\pm 15\%$ variation of the best-fit dipolar coupling (Table 4). Within the stated range of validity of eq 9, i.e., $D_{ik}\tau < 0.5$, the experimental points are observed to nearly all be within the $\pm 15\%$ bounds of the universal REAPDOR curve (eq 9). In this context, whereas in ref 34 experimental REAPDOR data are presented for well-isolated ${}^{13}C^{-67}Zn$ and ${}^{13}C^{-17}O$ spin pairs, van Wüllen et al. have shown by simulation that ²⁹Si/¹¹B REAPDOR curves for IS_2 systems are insensitive to geometry (for $\Delta S/S < 0.5$).⁵⁰ Moreover, ${}^{13}C-{}^{17}O$ REAPDOR buildup data for the tripeptide L-alanyl-alanyl-alanine in an antiparallel β -sheet arrangement have been shown to fit well to eq 9 using a single effective dipolar coupling.16

The fitted effective dipolar couplings in Table 4 are all observed to be significantly smaller than those determined from the geometrically optimized (CASTEP) crystal structures (see Table 2); this is a consequence of the partial ¹⁷O labeling. Rearranging eq 4, it follows that

$$p = \frac{D_{\text{rss},j}(\rho)^2}{D_{\text{rss},j}(\text{crystal})^2}$$
(10)

The proportions of ¹⁷O-labeled nuclei ρ in the glycine^{•2}HCl and uracil samples, as determined using eq 10 from the fitted



Figure 6. Experimental ¹⁵N/¹⁷O CP/REAPDOR curves, $\Delta S/S = (S - S_r)/S$, as a function of the dephasing time for (a) [²H(NH₃),1-¹³C, ¹⁵N, ¹⁷O₂]glycine ·²HCl and (b) [¹⁵N₂, ¹⁷O₂]uracil at various MAS frequencies. The best fits (assuming a single effective dipolar coupling, see Table 4) to eq 9 are shown as solid lines, with dashed lines corresponding to a ±15% variation of the best-fit dipolar coupling. Vertical dotted lines indicate the regions corresponding to $D_{jk}\tau < 0.5$ (see Table 4). The error bars for the signal intensity in experiments acquired with (S_r) and without (S) the dephasing pulse(s) were determined to be ±0.006 (glycine ·²HCl) and ±0.02 (uracil).

Table 4. $^{15}\text{N}{-}^{17}\text{O}$ Dipolar Coupling Parameters for Glycine $^{\text{2}}\text{HCl}$ and Uracil

site	$D_{jk} \pm$ 15%/Hz ^a	$D_{\rm rss,}/{\rm Hz}^b$	$ ho^c$	$ au$ ($D_{jk} au$ = 0.5)/ms
glycine• ² HCl N uracil N1	$\begin{array}{c} 88\pm13\\ 58\pm9 \end{array}$	123 173	0.37 - 0.68 0.08 - 0.15	5.6 8.6
uracil N3	114 ± 17	219	0.20-0.36	4.4

^{*a*} Determined from fits of the ${}^{15}N{-}{}^{17}O$ REAPDOR data in Figure 6 to eq 9. ^{*b*} As defined in eq 2, considering all N–O distances up to 6.0 Å in the geometrically optimized (CASTEP) crystal structures. ^{*c*} Determined using eq 10.

effective dipolar couplings for the 15 N/ 17 O CP/REAPDOR buildup data in Figure 6, are listed in Table 4. While both samples were prepared using [70%- 17 O]H₂O, the uracil sample is found to have a considerably lower degree of 17 O labeling. In this respect, it is to be noted that the uracil sample was prepared following the method described by Wu and co-workers, who estimated the degree of labeling in their sample as 14% at O2 and 24% at O4, starting with [40%- 17 O]H₂O.²⁵ This observation of non-uniform 17 O labeling is consistent with the lower ρ extracted for the N1 (0.11 ± 0.04) as compared to the N3 (0.26 ± 0.08) uracil REAPDOR curves: From inspection of the molecular structure, N1 has one close intramolecular proximity to O2, while N2 has close intramolecular proximities to O2 and O4.

6. Quantitative Measurement of Through-Bond *J* Couplings

J couplings are not normally resolved in ordinary onedimensional solid-state NMR spectra, since the observed line

⁽⁹³⁾ Vega, A. J. J. Magn. Reson. 1992, 96, 50.

⁽⁹⁴⁾ Grey, C. P.; Veeman, W. S.; Vega, A. J. J. Chem. Phys. 1993, 98, 7711.

⁽⁹⁵⁾ Grey, C. P.; Vega, A. J. J. Am. Chem. Soc. 1995, 117, 8232.

⁽⁹⁶⁾ Hughes, E.; Gullion, T.; Goldbourt, A.; Vega, S.; Vega, A. J. J. Magn. Reson. 2002, 156, 230.



Figure 7. ¹⁷O homonuclear spin–echo ($\tau/2 - \pi - \tau/2$) intensities for the (a,c) carbonyl (O1) and (b,d) hydroxyl (O2) ¹⁷O resonances in [²H(NH₃),1-¹³C,¹⁵N,¹⁷O₂]glycine ²HCl as a function of the evolution time together with best fits (solid lines, see Table 5) to (a,b) eq 11 and (c,d) eq 12. Experiments were carried out at 14.1 T (ν_0 (¹H) = 600 MHz) and a MAS frequency of 22.5 kHz. The error bars for the experimental intensities were determined to be ± 0.02 and are omitted for clarity.

Table 5. Parameters Extracted from the Fits of the ¹⁷O Glycine \cdot ²HCl Homonuclear Spin–Echo ($\tau/2 - \pi - \tau/2$) Intensities in Figure 7

data set	А	p ^a	<i>T_{2a}′/ms</i>	J ₀₀ /Hz	<i>T_{2b}</i> ′/ms	$[\varepsilon^2]^b$
$S_{\text{HOM}} \text{ O1 } (C=O)^c$	1.08 ± 0.01		39.2 ± 0.7			0.0037
$S_{\rm HOM}$ O2 (O ² H) ^c	0.91 ± 0.01		26.8 ± 0.6			0.0076
$S_{\text{HOM}} \text{ O1 } (\text{C=O})^{d,e}$	1.02 ± 0.01	0.34 ± 0.03	56.1 ± 4.2	8.8 ± 0.9	$56.1^{f} \pm 4.2$	0.0009
$S_{\text{HOM}} \text{ O2} (\text{O}^2\text{H})^{d,e}$	1.00 ± 0.02	0.73 ± 0.04	40.2 ± 3.9	4.5 ± 0.4	3.1 ± 1.1	0.0007

^{*a*} The proportion of ¹⁷O nuclei with an intramolecular ${}^{2}J_{OO}$ coupling, i.e., corresponding to the proportion of ¹⁷O-labeled nuclei. ^{*b*} $\varepsilon^{2} = \sum [I_{fin}(n) - I_{exp}(n)]^{2}/\sum I_{exp}(n)^{2}$. ^{*c*} Fit to eq 11. ^{*d*} Fit to eq 12. ^{*e*} One or more correlation coefficients are greater than 0.7 (see Supporting Information). ^{*f*} T_{2a} was set equal to T_{2b} (otherwise correlation coefficients of greater than 0.99 were obtained).

width is usually larger than the magnitude of the coupling. However, a simple $\tau/2 - \pi - \tau/2$ spin—echo MAS experiment refocuses evolution due to all terms that appear as offsets, e.g., magnetic field inhomogeneity or disorder, hence enabling the observation of *J* coupling modulation. The use of efficient heteronuclear decoupling gives transverse dephasing times, T_2' , which are usually much longer than the decay constants of the non-refocused NMR signal, T_2^* ,^{97,98} that characterize the freeinduction decay. Hence, it is possible to obtain refocused line widths ($\Delta' = 1/\pi T_2'$) that are sufficiently narrow that splittings due to *J* couplings can be observed. *J* modulation in MAS spin—echo experiments has been the subject of theoretical analysis.^{99–101} In particular, Duma et al. have provided a full

- (97) Cowans, B. A.; Grutzner, J. B. J. Magn. Reson. Ser. A 1993, 105, 10.
- (98) Lesage, A.; Bardet, M.; Emsley, L. J. Am. Chem. Soc. 1999, 121, 10987.
- (99) Kubo, A.; McDowell, C. A. J. Chem. Phys. 1990, 92, 7156.
- (100) Challoner, R.; Nakai, T.; McDowell, C. A. J. Chem. Phys. 1991, 94, 7038.
- (101) Duma, L.; Lai, W. C.; Carravetta, M.; Emsley, L.; Brown, S. P.; Levitt, M. H. *ChemPhysChem* **2004**, *5*, 815.

theoretical treatment of spin–echo *J* modulation for a homonuclear pair of spin I = 1/2 nuclei, supported by numerical simulations and experimental results on model systems.¹⁰¹ It was found that fitting the spin–echo modulation yields an accurate measurement of the *J* coupling constant, provided that, as is usually the case, rotational resonance conditions¹⁰² are avoided. This is found to be true even in the presence of large anisotropic interactions such as the chemical shift anisotropy and homonuclear dipolar coupling, which surprisingly act to stabilize the spin–echo modulation rather than to perturb it. Heteronuclear *J* couplings can also be measured using a heteronuclear spin–echo experiment, with the determination of the *J* couplings between a spin I = 1/2 and a half-integer quadrupolar nucleus having been recently reported for the ³¹P–²⁷Al^{37,38} and ³¹P–⁷¹Ga³⁹ spin pairs.

- (102) Raleigh, D. P.; Levitt, M. H.; Griffin, R. G. Chem. Phys. Lett. 1988, 146, 71.
- (103) Detken, A.; Hardy, E. H.; Ernst, M.; Meier, B. H. Chem. Phys. Lett. 2002, 356, 298.
- (104) Griffin, J. M.; Tripon, C.; Samoson, A.; Filip, C.; Brown, S. P. Magn. Reson. Chem. 2007, 45, S198.



Figure 8. (a-d) ${}^{17}O/{}^{13}C$ heteronuclear spin-echo ($\tau/2 - \pi - \tau/2$) intensities for the (a,c) carbonyl (O1) and (b,d) hydroxyl (O2) ${}^{17}O$ resonances in [${}^{2}H(NH_3)$,1- ${}^{13}C, {}^{15}N, {}^{17}O_2$]glycine ${}^{2}HCl$ as a function of the evolution time together with best fits to (a,b) eq 13 and (c,d) eq 14 (solid lines, see Table 6). Experiments were carried out at 14.1 T ($\nu_0({}^{1}H) = 600 \text{ MHz}$) and a MAS frequency of 22.5 kHz. The error bars for the experimental intensities were determined to be ± 0.02 and are omitted for clarity. (e,f) The quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$ for the (e) carbonyl (O1) and (f) hydroxyl (O2) ${}^{17}O$ resonances in [${}^{2}H(NH_3)$,1- ${}^{13}C, {}^{15}N, {}^{17}O_2$]glycine ${}^{2}HCl$ together with best fits to eq 15 (solid lines, see Table 6). $S_{\text{HET}}(\tau)$ and $S_{\text{HOM}}(\tau)$ refer to the ${}^{17}O/{}^{13}C$ heteronuclear and ${}^{17}O$ homonuclear spin-echo data presented in panels a and b of this figure and Figure 7, respectively. $S_Q(\tau)$ has been normalized such that $S_Q(\tau = 0) = 1.00$: experimentally the ratio of the integrated intensities $S_{\text{HET}}(\tau = 0)/S_{\text{HOM}}(\tau = 0)$ equaled 1.07 and 1.08 for the (e) carbonyl (O1) and (f) hydroxyl (O2) ${}^{17}O$ resonances, respectively.

Table 6. Parameters Extracted from the Fits of the	¹⁷ O- ¹³ C Glycine ² HCl Spir	in-Echo ($\tau/2-\pi-\tau/2$)) intensities in Figure 8
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data set	А	J _{CO} /Hz	<i>T_{2a}′/</i> ms	p ^a	J ₀₀ /Hz	<i>T</i> _{2b} ′/ms	$[\varepsilon^2]^b$
$\begin{array}{c} S_{\rm HET} \ {\rm O1} \ ({\rm C=O})^c \\ S_{\rm HET} \ {\rm O2} \ ({\rm O^2H})^c \\ S_{\rm HET} \ {\rm O1} \ ({\rm C=O})^{d.e} \\ S_{\rm HET} \ {\rm O2} \ ({\rm O^2H})^{d.e} \\ S_{\rm Q} \ {\rm O1} \ ({\rm C=O})^g \\ S_{\rm Q} \ {\rm O2} \ ({\rm O^2H})^g \end{array}$	$\begin{array}{c} 1.04 \pm 0.01 \\ 0.94 \pm 0.02 \\ 1.02 \pm 0.02 \\ 0.99 \pm 0.02 \\ 1.00 \pm 0.02 \\ 1.01 \pm 0.02 \end{array}$	$\begin{array}{c} 24.8 \pm 0.2 \\ 25.6 \pm 0.3 \\ 24.8 \pm 0.2 \\ 25.3 \pm 0.3 \\ 24.7 \pm 0.2 \\ 25.3 \pm 0.3 \end{array}$	$\begin{array}{c} 36.0 \pm 0.9 \\ 23.7 \pm 0.8 \\ 43.8 \pm 4.1 \\ 31.8 \pm 3.7 \\ 279^{h} \pm 70 \\ 231^{h} \pm 80 \end{array}$	$0.53 \pm 0.66 \\ 0.79 \pm 0.06$	5.5 ± 4.9 4.5 ± 0.9	$ \begin{array}{c} 43.8^{f} \pm 4.1 \\ 1.7 \pm 2 \end{array} $	$\begin{array}{c} 0.004 \\ 0.007 \\ 0.002 \\ 0.001 \\ 0.046 \\ 0.049 \end{array}$

^{*a*} The proportion of ¹⁷O nuclei with an intramolecular ${}^{2}J_{OO}$ coupling, i.e., corresponding to the proportion of ¹⁷O-labeled nuclei. ^{*b*} $\varepsilon^{2} = \sum [I_{fit}(n) - I_{exp}(n)]^{2} \sum I_{exp}(n)^{2}$. ^{*c*} Fit of the ¹⁷O/l³C heteronuclear spin—echo data to eq 13. ^{*d*} Fit of the ¹⁷O/l³C heteronuclear spin—echo data to eq 14. ^{*e*} One or more correlation coefficients are greater than 0.7 (see Supporting Information). ^{*f*} T_{2a} was set equal to T_{2b} (otherwise correlation coefficients of greater than 0.99 were obtained). ^{*s*} Fit of $S_{Q}(\tau) = S_{HET}(\tau)/S_{HOM}(\tau)$ to eq 15. ^{*h*} $1/\Delta T_{2}' = (1/T_{2 HET}') - (1/T_{2 HOM}')$.

6.1. Homonuclear ¹⁷**O** Spin–Echo: Glycine·²HCl. Experimental ¹⁷O homonuclear spin–echo ($\tau/2 - \pi - \tau/2$) intensities— acquired using the pulse sequence in Figure 1d (without the π

pulse on the nonobserve channel)—as a function of τ for the resolved C=O and C-O²H ¹⁷O sites of glycine [•]²HCl are shown in Figure 7. A faster dephasing is evident for the C-O²H ¹⁷O



Figure 9. (a) ¹³C homonuclear and (b) ¹³C/¹⁷O heteronuclear spin-echo $(\tau/2 - \pi - \tau/2)$ intensities for the carbonyl ¹³C resonance in [²H(NH₃),1-¹³C, ¹⁵N, ¹⁷O₂]glycine • ²HCl as a function of the evolution time together with best fits to eqs 11 and 17, respectively (solid lines, see Table 7). Experiments were carried out at 14.1 T ($\nu_0(^1\text{H}) = 600 \text{ MHz}$) and a MAS frequency of 22.5 kHz. The error bars for the experimental intensities were determined to be ± 0.002 and are omitted for clarity. (c) The quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/2$ the carbonyl ^{13}C resonance $S_{HOM}(\tau)$ for in $[^{2}H(NH_{3}),1-$ ¹³C, ¹⁵N, ¹⁷O₂]glycine ^{•2}HCl together with the best fit to eq 18 (solid line, see Table 7). $S_0(\tau)$ has been normalized such that $S_0(\tau = 0) = 1.00$: experimentally, the ratio of the integrated intensities $S_{\text{HET}}(\tau = 0)/S_{\text{HOM}}(\tau =$ 0) equaled 1.01.

site. The top row (Figure 7a,b) shows fits (solid lines, see Table 5) to a decaying exponential:

$$S(\tau) = A \exp(-\tau/T_{2a}') \tag{11}$$

where A is a normalization constant and T_{2a} is the "refocusable" transverse dephasing time.⁹⁸ While the fits in Figure 7a,b are good, a small systematic deviation of the data points from the decaying

exponential is observed, which suggests the presence of an additional modulation. A CASTEP⁴⁶ calculation (see section 2.3 and Supporting Information) predicts a 7.9 Hz ¹⁷O–¹⁷O two-bond intramolecular ²J_{OO} coupling for glycine [•]²HCl. It must be remembered, however, that ¹⁷O labeling is not complete; i.e., the sample contains molecules with none, one, or both of the oxygen sites ¹⁷O labeled. Therefore, for a specific resonance (i.e., O1 or O2), there are separate contributions from ¹⁷O nuclei with and without an intramolecular ²J_{OO} coupling. As shown in Figure 7c,d, better fits (solid lines, reduced ε^2 in Table 5) are obtained with the following function that takes into account such separate contributions from ¹⁷O nuclei with and without an intramolecular ²J_{OO} coupling:

$$S(\tau) = A[p \cos(\pi J \tau) \exp(-\tau / T_{2a}') + (1-p) \exp(-\tau / T_{2b}')]$$
(12)

A discussion of the comparison of the extracted *J* couplings to those determined by the CASTEP calculation will be given in the following section.

6.2. Heteronuclear ¹⁷O/¹³C Spin–Echo: Glycine ·²HCl. Experimental ¹⁷O/¹³C heteronuclear spin–echo ($\tau/2 - \pi - \tau/2$) intensities—acquired using the pulse sequence in Figure 1d—as a function of τ for the resolved C=O and C–O²H ¹⁷O sites of glycine ·²HCl are shown in Figure 8a,b. In both cases, clear *J* modulations are observed, and good fits (solid black lines, see Table 6) are obtained using eq 13.

$$S(\tau) = A\cos(\pi J \tau) \exp(-\tau / T_{2a}')$$
(13)

Note that the dephasing times for the O²H ¹⁷O resonance, 27 ± 1 ms for the ¹⁷O homonuclear and 24 ± 1 ms for the ¹⁷O/¹³C heteronuclear experiment, are slightly longer than that determined for ¹³CH-labeled L-alanine using XiX ¹H decoupling¹⁰³ ($\nu_1 = 100$ kHz) at 30 kHz MAS and 600 MHz ($T_2' = 20 \pm 2$ ms).¹⁰⁴

As was the case for the ¹⁷O homonuclear spin–echo data in Figure 7, slightly improved fits (see Figure 8c,d and Table 6) are obtained by considering the proportion of ¹⁷O nuclei with and without an intramolecular ${}^{2}J_{OO}$ coupling ($J_{1} = {}^{1}J_{CO}$, $J_{2} = {}^{2}J_{OO}$):

$$S(\tau) = A \cos(\pi J_1 \tau) [p \cos(\pi J_2 \tau) \exp(-\tau/T_{2a}') + (1-p) \exp(-\tau/T_{2b}')] \quad (14)$$

Tables 5 and 6 present four fits (correlation coefficients are given in the Supporting Information) that depend on the proportion of ¹⁷O nuclei, *p*, and the intramolecular ²J_{OO} coupling. The correlation coefficients involving *p* are lowest—specifically, the biggest correlation coefficient is that between *p* and T_{2b}' (0.27)—for the fit of $S_{\text{HET}}(\tau)$ for O2 (O²H) to eq 14, where the proportion of ¹⁷O nuclei is determined as 0.79 ± 0.06 (Table 6). In this context, it is to be noted that the sample was prepared using [70%-¹⁷O]H₂O. Considering ²J_{OO}, the lowest correlation coefficients are found for the fit of $S_{\text{HOM}}(\tau)$ for O1 (C=O) to eq 12 (*J*, *p* = -0.37 and *J*, $T_{2a}' =$ 0.67), where ²J_{OO} is determined as 8.8 ± 0.9 Hz. This is in good agreement with that predicted by the CASTEP⁴⁶ calculation (7.9 Hz) (see section 2.3 and Supporting Information).

Figure 8e,f shows the quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$, where $S_{\text{HET}}(\tau)$ and $S_{\text{HOM}}(\tau)$ refer to the ¹⁷O/¹³C heteronuclear and ¹⁷O homonuclear spin—echo data presented in Figure 8a,b and Figure 7, respectively. $S_O(\tau)$ is given as

$$S_{O}(\tau) = A \cos(\pi J \tau) \exp(-\tau / \Delta T_{2}')$$
(15)

where

$$1/\Delta T_2' = (1/T_{2 \text{ HET}}') - (1/\Delta T_{2 \text{ HOM}}')$$
(16)

Table 7. F	Parameters Extracted	from the Fits of the	¹³ C and ¹³ C-1	⁷ O Glycine · ² HCl Sp	in–Echo ($\tau/2-\pi-\tau/2$)	Intensities in Figure 9
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data set	А	<i>T_{2a}'/ms</i>	p ^a	<i>J</i> ₁ ^{<i>b</i>} /Hz	<i>T_{2b}</i> '/ms	$[\varepsilon^2]^c$
$S_{ ext{HOM}}^{d}$ $S_{ ext{HET}}^{e,f}$ $S_{ ext{Q}}^{f,g}$	$\begin{array}{c} 1.04 \pm 0.03 \\ 1.01 \pm 0.004 \\ 0.94 \pm 0.04 \end{array}$	$\begin{array}{c} 41.8 \pm 1.9 \\ 47.4 \pm 0.6 \\ 262^{h} \pm 149 \end{array}$	$0.23 \pm 0.01 \\ 0.15 \pm 0.02$	25.4 ± 2.7 26.2 ± 0.6	26.7 ± 0.7	0.0005 0.0004 0.0015

^{*a*} The proportion of ¹⁷O-labeled nuclei. ^{*b*} J₁ was set equal to J₂ (otherwise correlation coefficients of greater than 0.99 were obtained). ^{*c*} $\varepsilon^2 = \sum [I_{fit}(n) - I_{exp}(n)]^2 / \sum I_{exp}(n)^2$. ^{*d*} Fit to eq 11. ^{*e*} Fit to eq 17. ^{*f*} One or more correlation coefficients are greater than 0.7 (see Supporting Information). ^{*g*} Fit of S_Q(τ) = S_{HET}(τ)/S_{HOM}(τ) to eq 18. ^{*h*} 1/ $\Delta T_2' = (1/T_2 \text{ HeT}') - (1/T_2 \text{ HOM'}.)$



Figure 10. (a) ¹⁷O homonuclear (open diamonds) and ¹⁷O/¹⁵N heteronuclear (filled diamonds) spin–echo ($\tau/2 - \pi - \tau/2$) intensities for [¹⁵N₂,¹⁷O₂]uracil as a function of the evolution time together with best fits to eqs 11 and 19, respectively (solid lines, see Table 8). Experiments were carried out at 14.1 T ($\nu_0(^1\text{H}) = 600 \text{ MHz}$) and a MAS frequency of 20.0 kHz. The error bars for the experimental intensities were determined to be ± 0.04 and are omitted for clarity. (b) The quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$ for [¹⁵N₂,¹⁷O₂]uracil together with the best fit to eq 20 (solid line, see Table 8). $S_Q(\tau)$ has been normalized such that $S_Q(\tau = 0) = 1.00$. Experimentally the ratio of the integrated intensities $S_{\text{HET}}(\tau = 0)/S_{\text{HOM}}(\tau = 0)$ equaled 1.02.

Good fits (see Table 6) to eq 15 are observed in Figure 8e,f, with close agreement between the fitted *J* couplings of 24.7 \pm 0.2 and 25.3 \pm 0.3 Hz (the correlation coefficients involving *J* are less than 0.1) and those calculated using CASTEP⁴⁶ (see section 2.3 and Supporting Information), ¹*J*_{CO} = 24.9 and 27.5 Hz for the C=O and C-OH moieties, respectively. The large fitted values for $\Delta T_2'$ (279 and 231 ms) correspond to only a slight damping by the decaying exponential term and are consistent with the dephasing times obtained from fits of the heteronuclear spin-echo data to eq 13 (see Table 6) being only slightly shorter than those obtained from fits of the homonuclear spin-echo data to eq 11 (see Table 5): 36 cf. 39 ms (C=O) and 24 cf. 27 ms (O²H).

6.3. Homonuclear ¹³C and Heteronuclear ¹³C/¹⁷O Spin– Echo: Glycine ²HCl. ¹³C homonuclear and ¹³C/¹⁷O heteronuclear spin–echo ($\tau/2 - \pi - \tau/2$) intensities as a function of τ for glycine ²HCl (¹³C labeled only at the carboxylic acid carbon)

Table 8.	Parameters	Extracted	from the	Fits of the	¹⁷ O and	
17O-15N	Uracil Spin-	Echo (r/2	$-\pi - \tau/2$)	Intensities	in Figure	10

data set	А	<i>T_{2a}′/ms</i>	p ^a	^{2h} J _{NO} ^b /Hz	$[\varepsilon^2]^c$
$S_{\text{HOM}}^{d,e}$	0.87 ± 0.03	102 ± 7	0.66 + 0.14	50106	0.0075
S_{HET}	0.96 ± 0.04 1.01 ± 0.05	$62^{s} \pm 10$ $550^{i} \pm 1479$	0.66 ± 0.14 0.85 ± 0.20	5.8 ± 0.6 5.1 ± 0.6	0.0026

^{*a*} The relative contribution of O4 ¹⁷O nuclei to the ¹⁷O resonance (overlapping O2 and O4). ^{*b*} J₁ was set equal to J₂ (otherwise correlation coefficients of greater than 0.99 were obtained). ^{*c*} $\varepsilon^2 = \sum [I_{\rm fit}(n) - I_{\rm exp}(n)]^2 / \Sigma I_{\rm exp}(n)^2$. ^{*d*} Fit to eq 11. ^{*e*} One or more correlation coefficients are greater than 0.7 (see Supporting Information). ^{*f*} Fit to eq 19. ^{*g*} T'_{2a}was set equal to T'_{2b}(otherwise correlation coefficients of greater than 0.99 were obtained). ^{*h*} Fit of $S_Q(\tau) = S_{\rm HET}(\tau)/S_{\rm HOM}(\tau)$ to eq 20. ^{*i*} 1/ $\Delta T_2' = (1/T_2 _{\rm HET}') - (1/T_2 _{\rm HOM}')$.

acquired using the pulse sequence in Figure 1e (without and with the ¹⁷O π pulse) are shown in Figure 9a,b, respectively. The ¹³C homonuclear spin-echo data (Figure 9a) are well fit (see Table 7) to the simple decaying exponential function of eq 11.

Unlike for the ¹⁷O/¹³C heteronuclear spin—echo data in Figure 8a,b, zero-crossings are not observed for the ¹³C/¹⁷O heteronuclear spin—echo data (see Figure 9b). In this context, the partial ¹⁷O enrichment of the oxygen sites must be remembered, with there being a proportion $(1 - p)^2$ of the ¹³C nuclei having no one-bond *J* coupling to ¹⁷O—a situation similar to the ¹³C spin—echo data presented in ref 105 for a partially enriched cellulose sample. The experimental data were fit (solid line in Figure 9b) using the function

$$S(\tau) = A[\exp(-\tau/T_{2a}')\{p^{2}\cos(\pi J_{1}\tau)\cos(\pi J_{2}\tau) + p(1-p)\cos(\pi J_{1}\tau) + p(1-p)\cos(\pi J_{2}\tau)\} + \exp(-\tau/T_{2b}')(1-p)^{2}] \quad (17)$$

where *p* is the probability that one oxygen site is ¹⁷O labeled (this is assumed to be the same for the two oxygen sites), and separate dephasing times are considered for ¹³C with and without a ¹⁷O neighbor(s). A good fit (setting $J_1 = J_2$) of the experimental data to eq 17 is evident from Figure 9b; however, high correlation coefficients involving J (J, p = -0.65; J, $T_{2a}' = -0.61$; J, $T_{2b}' = 0.40$) and p (p, J = -0.65; p, $T_{2a}' = 0.87$; p, $T_{2b}' = -0.66$) are observed.

Figure 9c shows the quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$, where $S_{\text{HET}}(\tau)$ and $S_{\text{HOM}}(\tau)$ refer to the ¹³C/¹⁷O heteronuclear and ¹³C homonuclear spin—echo data presented in Figure 9a,b. $S_Q(\tau)$ is given as

$$S_{Q}(\tau) = A \{ \exp(-\tau/\Delta T_{2}') [p^{2} \cos(\pi J_{1}\tau) \cos(\pi J_{2}\tau) + p(1-p) \cos(\pi J_{1}\tau) + p(1-p) \cos(\pi J_{2}\tau)] + (1-p)^{2} \}$$
(18)

Note that the decaying exponential term corresponding to the difference ($\Delta T_2'$ is defined in eq 16) between the dephasing times for the heteronuclear and homonuclear spin-echo is omitted for the proportion of ¹³C without a directly bonded ¹⁷O nucleus because the ¹⁷O π pulse is not expected to change the dephasing time for this proportion of ¹³C without a directly



Figure 11. (a,b) ¹⁵N homonuclear (open diamonds) and ¹⁵N/¹⁷O heteronuclear (filled diamonds) spin–echo ($\tau/2 - \pi - \tau/2$) intensities for the (a) N1 and (b) N3 resonances of [¹⁵N₂,¹⁷O₂]uracil as a function of the evolution time. Experiments were carried out at 14.1 T ($\nu_0(^{1}\text{H}) = 600 \text{ MHz}$) and a MAS frequency of 20.0 kHz. The error bars for the experimental intensities were determined to be ± 0.0008 and are omitted for clarity. (c,d) The quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$ for the (c) N1 and (d) N3 resonances of [¹⁵N₂,¹⁷O₂]uracil together with the best fits (solid line, see Table 9) to eq 21. $S_Q(\tau)$ has been normalized such that $S_Q(\tau = 0) = 1.00$. Experimentally the ratio of the integrated intensities $S_{\text{HET}}(\tau = 0)/S_{\text{HOM}}(\tau = 0)$ equaled 1.001 and 0.999 for the (c) N1 and (d) N3 resonances, respectively.

Table 9.	Parameters	Extracted	from the	Fits of the	¹⁵ N and	¹⁵ N-1/C) Uracil Spir	ı−Echo (τ/	/2-π-τ/2)) Data in I	Figures 1	1 and 1	2
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data set	А	p ^a	J _{NO} /Hz	J _{NN} /Hz	<i>T_{2a}′/ms</i>	<i>T</i> _{2b} ′/ms	$[\varepsilon^2]^b$
$S_Q N1 - O4^{c,d}$	1.00 ± 0.01	0.22 ± 0.01	6.7 ± 0.4		$294^e \pm 38$		0.0003
$S_Q N3 - O4^{c,d}$	1.00 ± 0.01	0.27 ± 0.03	4.8 ± 0.5		$585^{e} \pm 98$		0.0003
$S_{\text{HET}} \text{N1}-\text{O4}^{d,f}$	0.93 ± 0.01	0.51 ± 0.15	0.0 ± 67	2.7 ± 0.1	121 ± 4	33.7 ± 4.6	0.0005
$S_{\rm HET}$ N3 $-$ O4 ^{d,f}	0.94 ± 0.01	0.69 ± 0.07	0.0 ± 1775	2.6 ± 0.1	110 ± 2	25.4 ± 3.1	0.0003

^{*a*} The proportion of ¹⁵N nuclei with a *J* coupling to an ¹⁷O nucleus, corresponding to the degree of ¹⁷O labeling at the O4 site. ^{*b*} $\varepsilon^2 = \sum [I_{fit}(n) - I_{exp}(n)]^2 / \sum I_{exp}(n)^2$. ^{*c*} Fit of $S_Q(\tau) = S_{HET}(\tau)/S_{HOM}(\tau)$ to eq 21; *J* corresponds to ^{2h} $J_{N,O}$. ^{*d*} One or more correlation coefficients are greater than 0.7 (see Supporting Information). ^{*e*} $1/\Delta T_2' = (1/T_2 + HET') - (1/T_2 + HOM')$. ^{*f*} Fit to eq 14.

bonded ¹⁷O nucleus. A good fit (setting $J_1 = J_2$, see Table 7) to eq 18 is observed in Figure 9c. The correlation coefficients involving J are small (magnitude 0.04–0.12) for the fit of the $S_Q(\tau)$ data—this is in contrast to the fit of the heteronuclear spin—echo data in Figure 9b—and close agreement between the fitted J coupling (26.7 ± 0.6 Hz) and the average of those determined by CASTEP (${}^{1}J_{CO} = 24.9$ and 27.5 Hz for the C=O and C—OH moieties, respectively) is observed. In contrast, high correlation coefficients involving p (p, $\Delta T_2' = -0.76$; p, A = 0.93) mean that the degree of 17 O labeling cannot be reliably determined from this fit.

6.4. Homonuclear ¹⁷O and Heteronuclear ¹⁷O/¹⁵N Spin– Echo: Uracil. In the uracil molecule, there are two distinct nitrogen and oxygen sites, with the crystal structure (Figure 2b) of uracil revealing that only one of the two distinct oxygen atoms participates in NH···O hydrogen-bonding: oxygen O4 forms two intermolecular NH···O hydrogen bonds to N1 and N3. In the ¹⁷O MAS spectrum of [¹⁵N₂,¹⁷O₂]uracil shown in Figure 3b, the second-order quadrupolar broadened line shapes for the two distinct ¹⁷O resonances are unresolved from each other. In a spin–echo analysis, it is thus necessary to consider the contribution from 17 O nuclei with and without hydrogen-bondmediated *J* couplings to N1 and N3.

Figure 10a compares homonuclear ¹⁷O (open diamonds) and heteronuclear ¹⁷O/¹⁵N (filled diamonds) spin—echo ($\tau/2 - \pi - \tau/2$) data obtained using the pulse sequence in Figure 1d (without and with the ¹⁵N π pulse). Best fits (see Table 8) are shown as solid lines for the homonuclear spin—echo data fit to the simple decaying exponential function of eq 11 and the heteronuclear spin—echo data (for J_1 set equal to J_2 and T_{2a} ' set equal to T_{2b} ') fit to

$$S(\tau) = A[p \cos(\pi J_1 \tau) \cos(\pi J_2 \tau) \exp(-\tau / T_{2a}') + (1-p) \exp(-\tau / T_{2b}')] \quad (19)$$

Figure 10b shows the quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$, where $S_{\text{HET}}(\tau)$ and $S_{\text{HOM}}(\tau)$ refer to the ¹⁷O/¹⁵N heteronuclear and ¹⁷O homonuclear spin—echo data presented in Figure 10a. $S_Q(\tau)$ is given as

$$S_{Q}(\tau) = A\{p \cos(\pi J_{1}\tau) \cos(\pi J_{2}\tau) \exp(-\tau/\Delta T_{2}') + (1-p)\}$$
(20)



Figure 12. ¹⁵N/¹⁷O heteronuclear spin–echo ($\tau/2 - \pi - \tau/2$) intensities for the (a) N1 and (b) N3 ¹⁵N sites of [¹⁵N₂,¹⁷O₂]uracil as a function of the evolution time together with the best fits (black curves) to eq 14 (see Table 9). The error bars for the experimental ¹⁵N data points of both sites were determined to be ±0.003 and are omitted for clarity (16 transients were coadded for each τ increment). Expansions for $\tau = 160-250$ ms are shown as insets. Experiments were carried out at 14.1 T (ν_0 (¹H) = 600 MHz) and a MAS frequency of 20.0 kHz.

A good fit (for J_1 set equal to J_2 , see Table 8) to eq 20 is observed in Figure 10b. The ^{2h} J_{NO} couplings as determined from the fits of $S_{\text{HET}}(\tau)$ and $S_Q(\tau)$, namely 5.8 ± 0.6 and 5.1 ± 0.6 Hz, respectively, are in agreement with each other. Smaller correlation coefficients involving J are found for the fit of $S_Q(\tau)$, although those for $S_{\text{HET}}(\tau)$ are still reasonable (J, p = 0.52; J, $T_{2a'} = -0.41$; J, A = 0.37 for $S_{\text{HET}}(\tau)$; and J, p = -0.25; J, $\Delta T_2' = -0.01$; J, A = 0.15 for $S_Q(\tau)$). The experimental average ^{2h}J_{NO} couplings are in good agreement with those determined by a CASTEP⁴⁶ calculation (see section 2.3 and Supporting Information), namely ^{2h}J_{N1,O4} = 6.1 and ^{2h}J_{N3,O4} = 4.6 Hz. As shown in the Supporting Information, all other ¹⁵N-¹⁷O J couplings (both inter- and intramolecular) are calculated to be of magnitude 0.5 Hz or less. Note that CASTEP calculations have recently been shown to provide excellent accuracy for hydrogen-bond-mediated ^{2h}J_{NN} couplings¹⁰⁶ determined experimentally using ¹⁵N spin—echo MAS experiments.^{40,107}

mentally using ¹⁵N spin–echo MAS experiments.^{40,107} 6.5. Homonuclear ¹⁵N and Heteronuclear ¹⁵N/¹⁷O Spin–Echo: Uracil. Figure 11a,b compares homonuclear ¹⁵N (open diamonds) and heteronuclear ¹⁵N/¹⁷O (filled diamonds) spin–echo ($\tau/2 - \pi - \tau/2$) data obtained using the pulse sequence in Figure 1e (without and with the ¹⁷O π pulse) for the N1 and N3 resonances. In both cases, there is a small yet significant difference between the homonuclear and heteronuclear spin–echo data. Figure 11c,d shows the quotient $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$ for the N1 and N3 resonances, where $S_{\text{HET}}(\tau)$ and $S_{\text{HOM}}(\tau)$ refer to the ¹⁵N/¹⁷O heteronuclear and ¹⁵N homonuclear spin–echo data presented in Figure 11a,b. $S_O(\tau)$ is given as

$$S_{0}(\tau) = A\{p \cos(\pi J \tau) \exp(-\tau / \Delta T_{2}') + (1-p)\}$$
(21)

where *p* corresponds to the proportion of ¹⁵N nuclei with a *J* coupling to an ¹⁷O nucleus. Good fits (see Table 9) to eq 21 are observed in Figure 11c,d. Although there is a very high correlation between *p* and *J* (the *p*, *J* correlation coefficient equals -0.98 and -0.99 for N1 and N3, respectively), the fitted *J* couplings of 6.7 ± 0.4 Hz (N1) and 4.8 ± 0.5 Hz (N3) are in remarkably good agreement with those determined by the CASTEP⁴⁶ calculation, namely ^{2h}J_{N1,O4} = 6.1 and ^{2h}J_{N3,O4} = 4.6 Hz.

Figure 12 presents ${}^{15}N/{}^{17}O$ heteronuclear spin-echo ($\tau/$ $2-\pi-\tau/2$) data for the N1 and N3 uracil resonances for a spin-echo duration up to 250 ms (obtained experimentally separately from the data presented in Figure 11). The CASTEP⁴⁶ calculation (see section 2.3 and Supporting Information) predicts an intramolecular $J_{\rm NN}$ coupling in uracil, ${}^{2}J_{\rm N1,N3} = 2.7$ Hz. This is consistent with the observation of zero-crossings at long spin-echo durations (~200 ms) and the good fits (see Table 9) to eq 14 (where $J_1 = {}^2J_{NN}$, $J_2 = {}^{2h}J_{NO}$). Specifically, the correlation coefficients involving ${}^{2}J_{NN}$ are reasonable (maximum magnitude 0.59 and 0.62 for the N1 and N3 data, respectively), and the experimental values of ${}^{2}J_{\rm NN} = 2.7 \pm 0.1$ and 2.6 ± 0.1 Hz are in perfect agreement with the calculated value of ${}^{2}J_{N1,N3}$ = 2.7 Hz. Table 9 shows that it is, however, not possible to determine ${}^{2h}J_{NO}$ from the fit to the heteronuclear spin-echo data. In this respect, note that the modulation under the homonuclear ${}^{2}J_{\rm N1,N3}$ coupling will be present for both the ${}^{15}\rm N$ homonuclear and ¹⁵N/¹⁷O heteronuclear spin-echo experiments. It is removed by taking the quotient $S_O(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$, hence allowing the observation in Figure 11c,d of the modulation due to the proportion of ¹⁵N with a hydrogen-bond-mediated J coupling to a ¹⁷O nucleus.

7. Conclusions

The feasibility of heteronuclear solid-state MAS NMR experiments for probing ${}^{15}N{}-{}^{17}O$ dipolar and J couplings in two isotopically labeled model compounds, glycine · ²HCl and uracil, has been demonstrated. Specifically, the R³-HMQC experiment yields two-dimensional ¹⁵N-¹⁷O correlations for dipolar coupled ¹⁵N and ¹⁷O nuclei; for glycine^{•2}HCl, the intensity of the resolved peaks for the C=O and O²H ¹⁷O resonances corresponds to the relative magnitude of the respective ¹⁵N-¹⁷O dipolar couplings. Fits of the initial buildup (ΔS /S < 0.2) in ${}^{17}\text{O}-{}^{15}\text{N}$ REDOR curves for glycine ${}^{2}\text{HCl}$ yield effective dipolar couplings in agreement with $(\pm 20\%)$ the rootsum-squared dipolar couplings⁴⁹ determined from the crystal structure. Experimental ¹⁵N-¹⁷O REAPDOR curves for the ¹⁵N resonance in glycine^{•2}HCl and the two distinct ¹⁵N resonances in uracil are well fit to the universal curve presented by Goldbourt et al.34

The first (to the best of our knowledge) solid-state NMR determination of ${}^{13}\text{C}{-}^{17}\text{O}$, ${}^{17}\text{O}{-}^{17}\text{O}$, and ${}^{15}\text{N}{-}^{17}\text{O}$ *J* couplings is reported. Heteronuclear ${}^{13}\text{C}{-}^{17}\text{O}$ and ${}^{15}\text{N}{-}^{17}\text{O}$ *J* couplings are experimentally determined from fits of the quotient of the integrated intensity obtained in a heteronuclear and a homonuclear spin–echo experiment, $S_Q(\tau) = S_{\text{HET}}(\tau)/S_{\text{HOM}}(\tau)$. Importantly, for $S_Q(\tau)$, there is only a small damping of the signal

⁽¹⁰⁵⁾ Brown, S. P.; Emsley, L. J. Magn. Reson. 2004, 171, 43.

⁽¹⁰⁶⁾ Joyce, S. A.; Yates, J. R.; Pickard, C. J.; Brown, S. P. J. Am. Chem. Soc. 2008, 130, 12663.

⁽¹⁰⁷⁾ Brown, S. P.; Perez-Torralba, M.; Sanz, D.; Claramunt, R. M.; Emsley, L. Chem. Commun. 2002, 1852.

that is fit to a decaying exponential corresponding to the *difference* in the dephasing times for the heteronuclear and homonuclear spin-echo experiments. Excellent agreement is observed between the experimentally determined J couplings and those calculated using the first-principles CASTEP code.⁴⁶

To the best of our knowledge, this is the first demonstration of ${}^{15}N{-}^{17}O$ solid-state NMR experiments. ${}^{17}O$ solid-state NMR is being increasingly employed as a probe of biological molecules. ${}^{6,24,25,52,108-110}$ Given the importance of NH····O hydrogen-bonding, e.g., in proteins and nucleic acids, there is evidently much potential to further apply the ${}^{15}N{-}^{17}O$ solid-state NMR experiments demonstrated in this paper, so as to

- (109) Lemaitre, V.; de Planque, M. R. R.; Howes, A. P.; Smith, M. E.; Dupree, R.; Watts, A. J. Am. Chem. Soc. 2004, 126, 15320.
- (110) Kwan, I. C. M.; Mo, X.; Wu, G. J. Am. Chem. Soc. 2007, 129, 2398.

unambiguously identify specific structure-determining hydrogenbonding interactions.

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Supporting Information Available: Correlation coefficients for fits of spin–echo data, spin–echo spectra obtained for $\tau = 0$, full listing of calculated *J* couplings, complete ref 45, and geometrically optimized (CASTEP) crystal structures, as used for the calculation of the NMR *J* couplings. This material is available free of charge via the Internet at http://pubs.acs.org.

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⁽¹⁰⁸⁾ Wong, A.; Howes, A. P.; Pike, K. J.; Lemaitre, V.; Watts, A.; Anupold, T.; Past, J.; Samoson, A.; Dupree, R.; Smith, M. E. J. Am. Chem. Soc. 2006, 128, 7744.